



The National Nanotechnology Initiative presents

us-eu *bridging nanoEHS research efforts*

A Joint US-EU Workshop

March 10 - 11, 2011

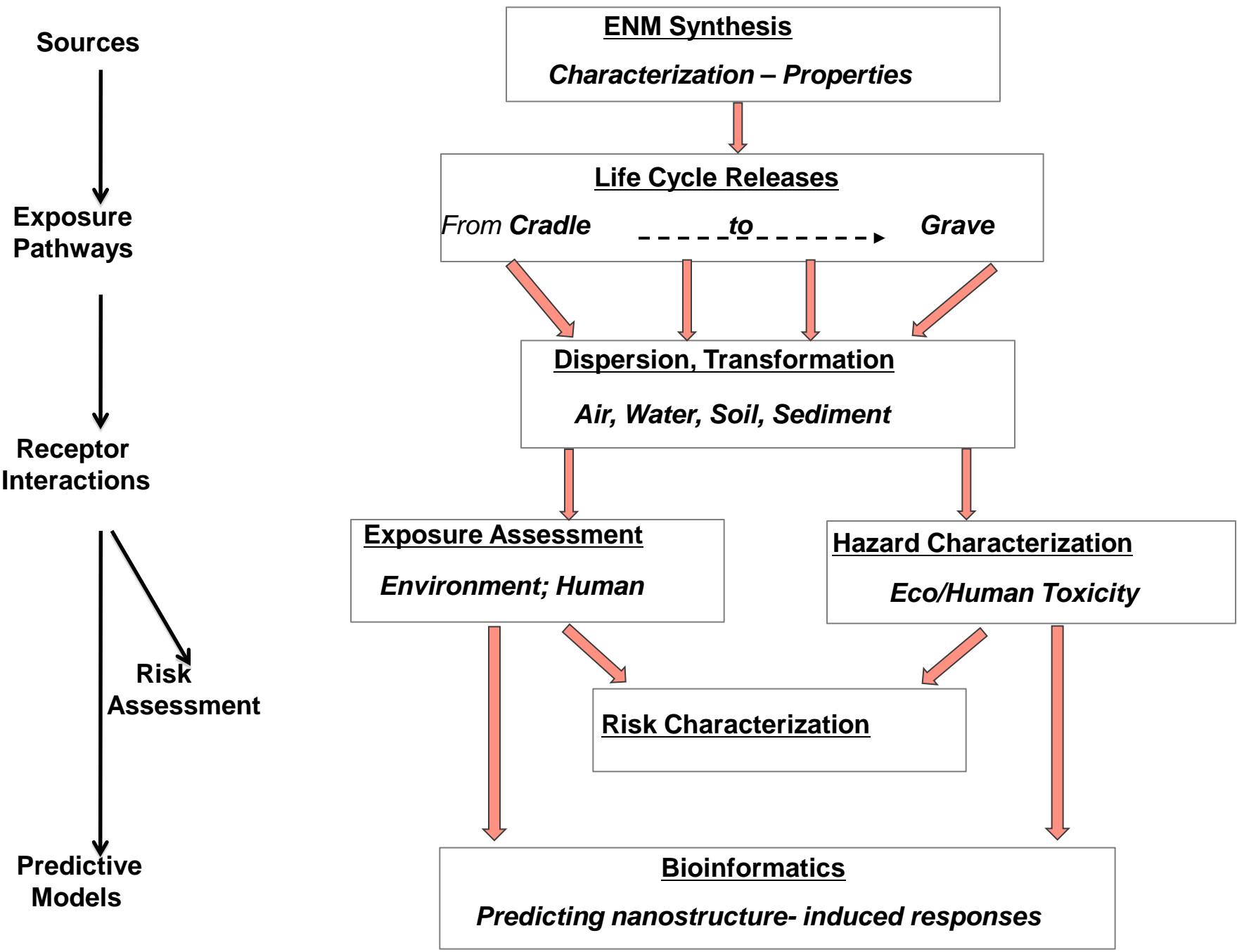
Washington, DC

In Vitro –In Vivo Correlations of Dose- and Response-Metrics:
Concepts for OEL Extrapolation

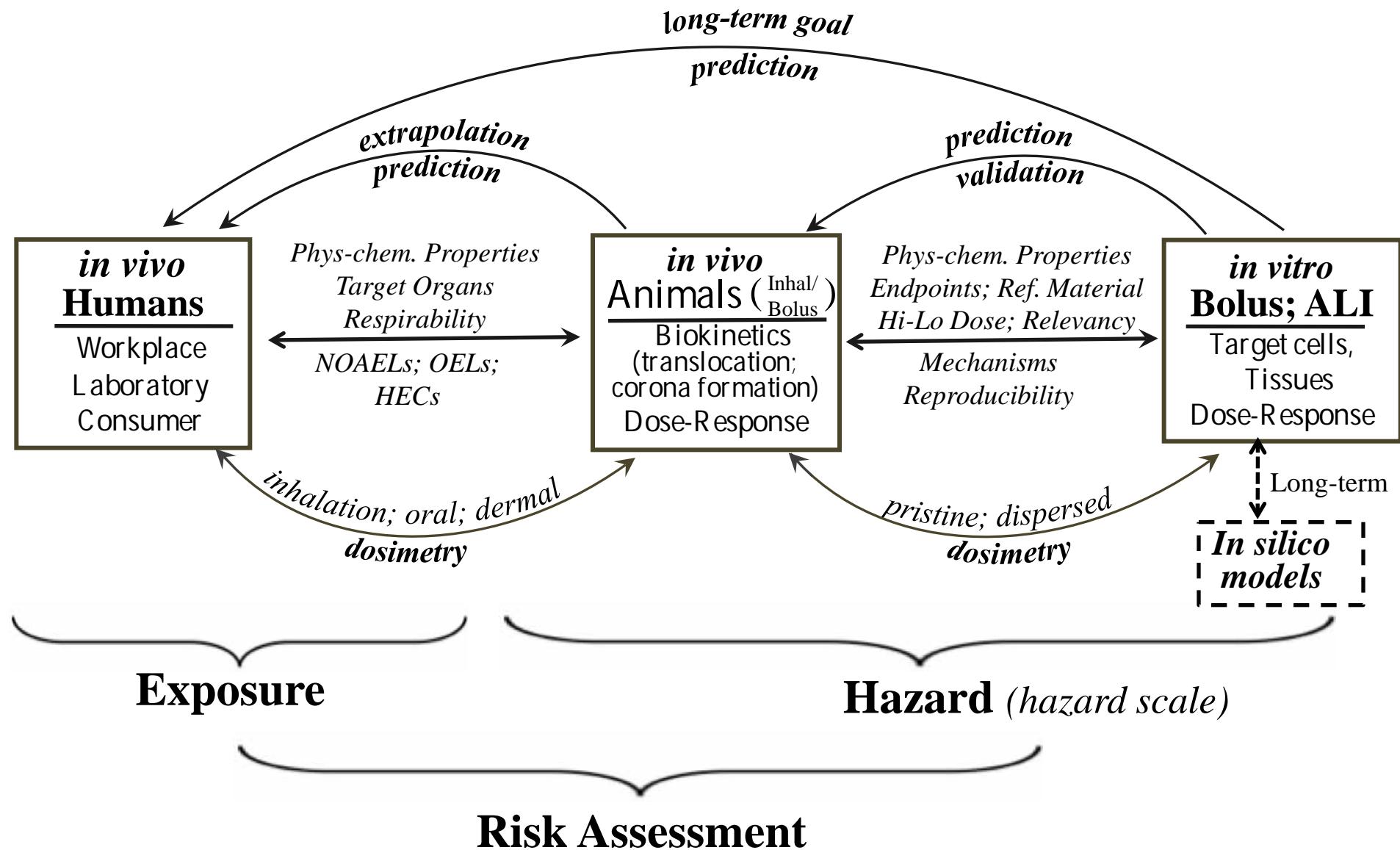
Günter Oberdörster
University of Rochester
March 11, 2011



Data Needs for ENM Induced Environmental and Health Effects and Associated Risks

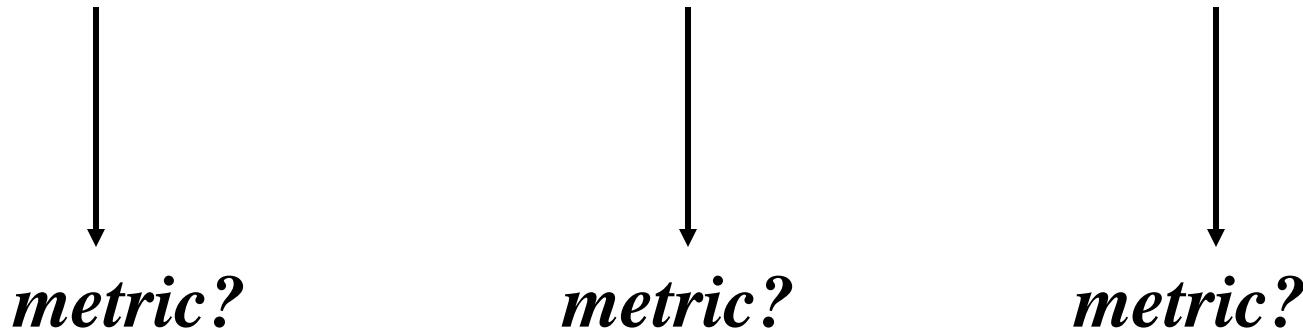


Concepts of Nanomaterial Toxicity Testing: *Risk Assessment as Function of Exposure and Hazard*



“Best” Dosemetric?

Exposure >>>>> Dose >>>>> Response



Mass/Number/surface Area Correlations for Selected NPs

Particle Type	Diameter nm	Density g/cm ³	Spc. Srf. Area m ² /g	Specific # number/g	Airborne Conc. of 100 µg/m ³	
					Surface cm ² /m ³	Number #/m ³
Pt	50	21.09	5.69	7.24x10 ¹⁴	5.69	7.24x10 ¹⁰
Gold	50	19.3	6.22	7.91x10 ¹⁴	6.22	7.92x10 ¹⁰
Ag	50	7.2	11.43	1.46x10 ¹⁵	11.43	1.46x10 ¹¹
Cu	50	8.9	13.48	1.72x10 ¹⁵	13.48	1.72x10 ¹¹
Al	50	2.7	44.44	5.65x10 ¹⁵	44.44	5.65x10 ¹¹
TiO ₂ (R)	50	4.23	28.37	3.61x10 ¹⁵	28.37	3.61x10 ¹¹
TiO ₂ (A)	50	3.9	30.77	3.92x10 ¹⁵	30.77	3.92x10 ¹¹
C	50	2.26	53.10	6.76x10 ¹⁵	53.10	6.76x10 ¹¹
Polystyrene	50	1.05	114.3	1.46x10 ¹⁶	114.3	1.46x10 ¹²

Usual Physical Dose-Metrics for NPs that Correlate with Biol./Toxicol. Effects:

Mass
Number
Surface Area
(Volume)

}

correlation between these should be part of NP characterization

BET Surface Area:

Which one?

N; Kr; Ar; others?

Need for standardization: Most common is use of nitrogen.

Most desirable: Measure of bioavailable SA

Also: *BET equivalent particle size:*

to characterize/estimate agglomeration/aggregation

Physico-Chemical Properties of Investigated Nanoparticles

Particle	Origin	Primary Particle Size (nm) (a)	Crystal Phase	Specific Surface Area (m ² /gm)	BET Equivalent Diameter (nm)	Hydrodynamic Diameter (nm) (b)	Zeta Potential (mV) (c)	Agglomm/ Aggregation Degree
Elemental Carbon (EC)	Electric Spark Generated (Rochester)	41	Amorphous	767.9	3.4	204	-50.5	High
TiO ₂ (F)	Fischer Scientific	250	Anatase	8.0	195	1287	-14.7	Medium
TiO ₂ (M)	Millenium Chemical Corp.	~20	Anatase	86.1	18.3	1608	14	High
TiO ₂ (D)	Evonik Chemicals	~25	80% anatase/ 20% rutile	57.4	27	576	27.3	Medium
Copper-40	Nanotechnologies	40	FCC crystal	30.6	21.9	850	-0.6	Medium
Silver-35	Nanotechnologies	35	FCC crystal	21.0	27.3	483	-47.0	Medium
Au ₅₀	Ted Pella (Ca)	50	FCC crystal	6	51.8	93	-33.8	Low
PS-NH ₃	Bangs Laboratories	65	Amorphous	88	65	72	83.0	Very low

(a) Size as reported by manufacturer;

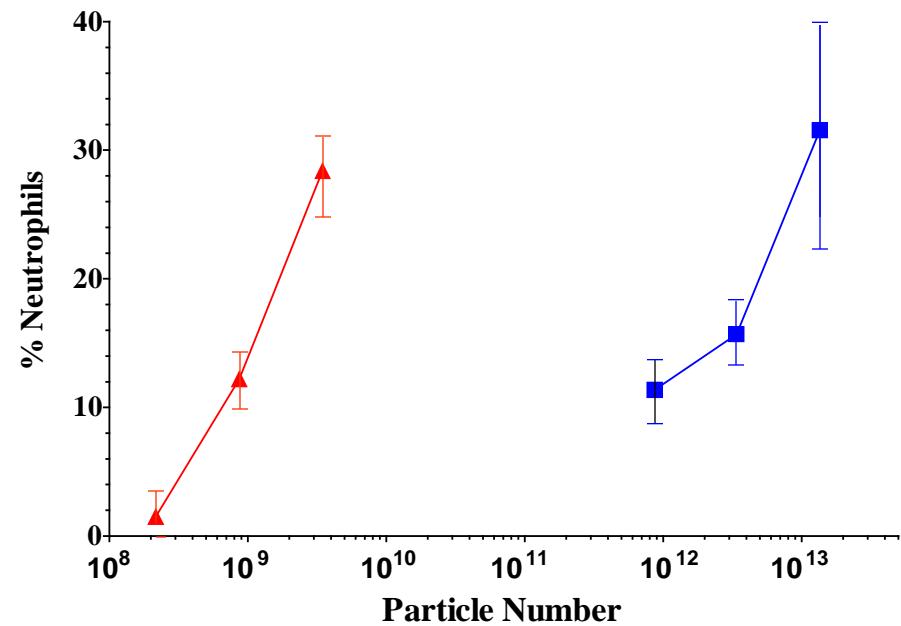
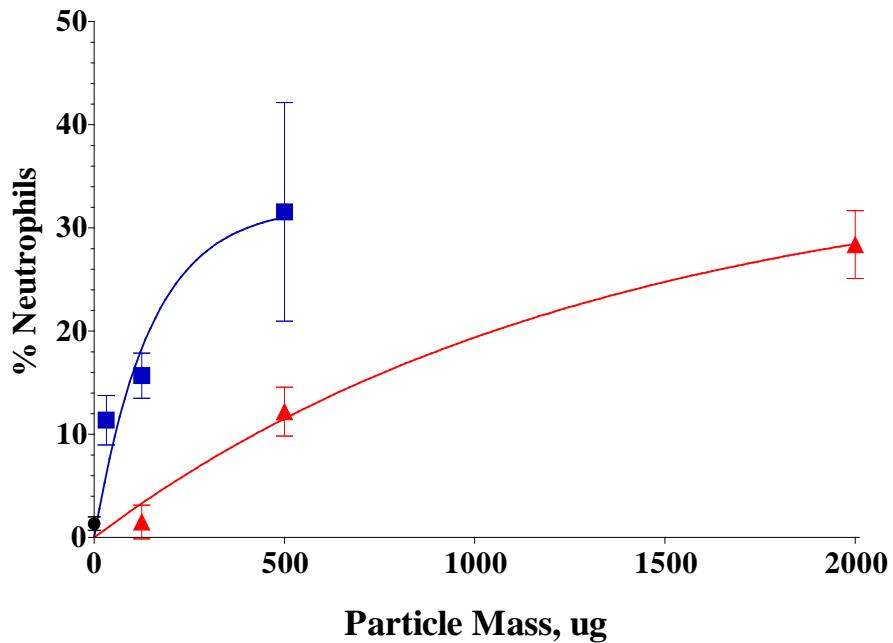
(b) measured by dynamic light scattering (DLS) in water;

(c) measured by electrophoretic light scattering (ELS) in water.

Which Dose-Metric?

Percent of Neutrophils in Lung Lavage 24 hrs after
Intratrachael Dosing of Ultrafine and Fine TiO_2 in Rats

fine TiO_2 (200nm)
ultrafine TiO_2 (25nm)
saline



Particle Mass

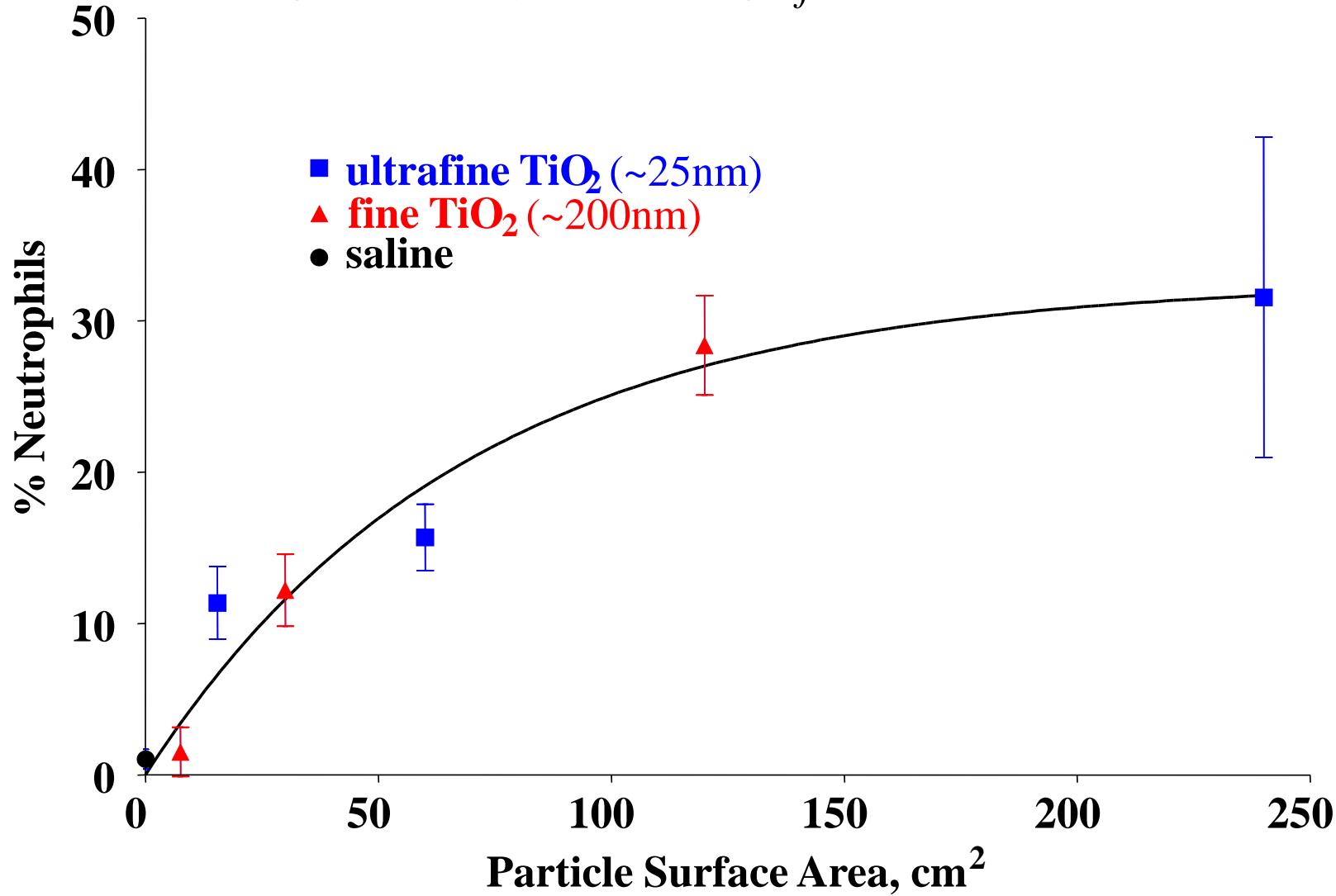
vs

Particle Number

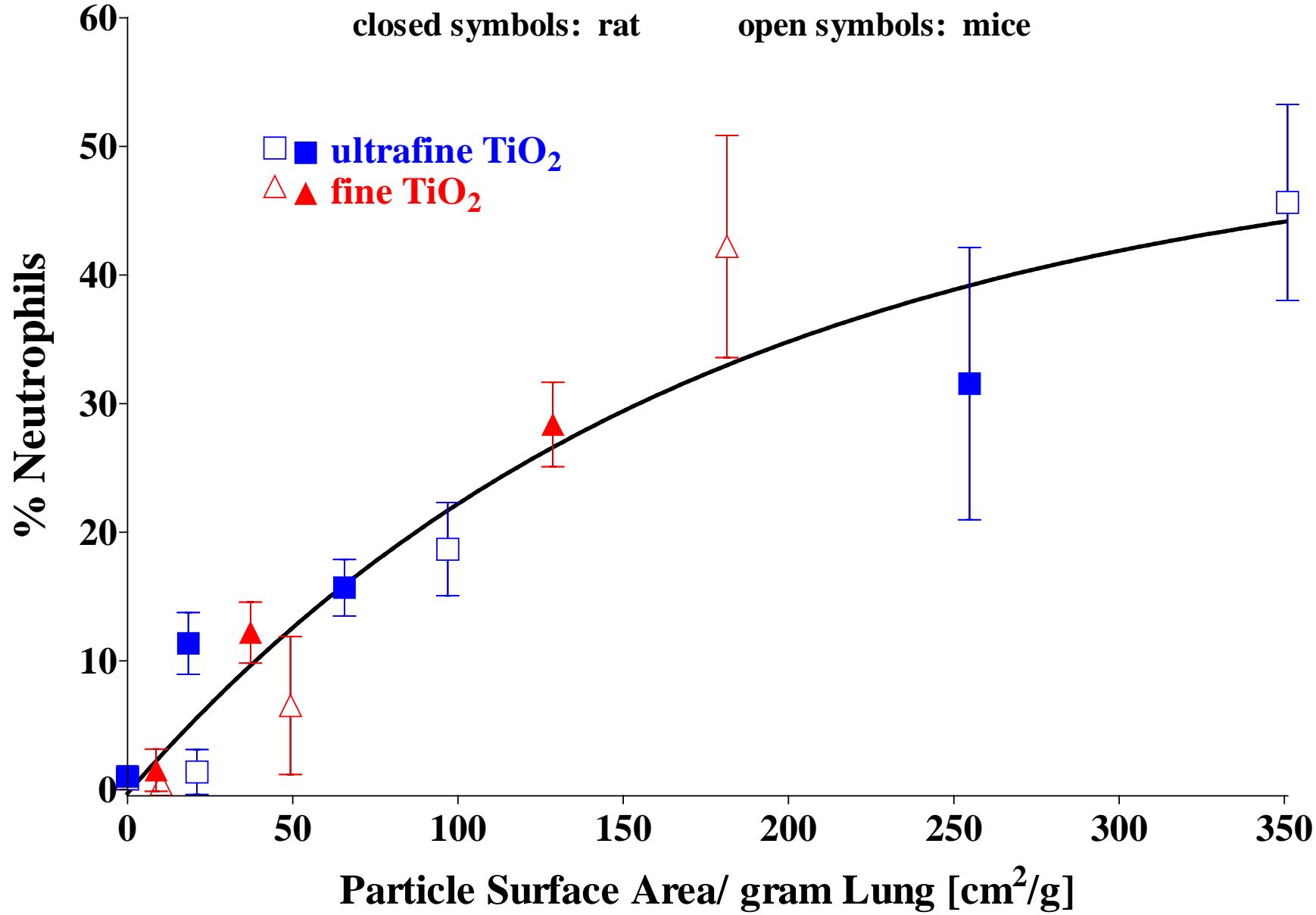
Which Dose-Metric?

Percent of Neutrophils in BAL 24 hrs after Instillation of TiQin Rats

Correlation with Particle Surface Area



Percent of Neutrophils in BAL 24 hrs after Instillation



**In addition: “Chemical” Dose-Metric,
e.g., ROS inducing potential:**

DCFH-DA (*2’-7’ dichlorofluorescin-diacetate*) assay

FRAS (*ferric reducing ability of serum*) assay

Vit C assay

others...

(as screening tool for categorization of NPs based on reactivity [Bello et al., 2009; Rushton et al., 2010])

ROS-Inducing Capacity of NanoTiO₂ depends on Crystalline State:

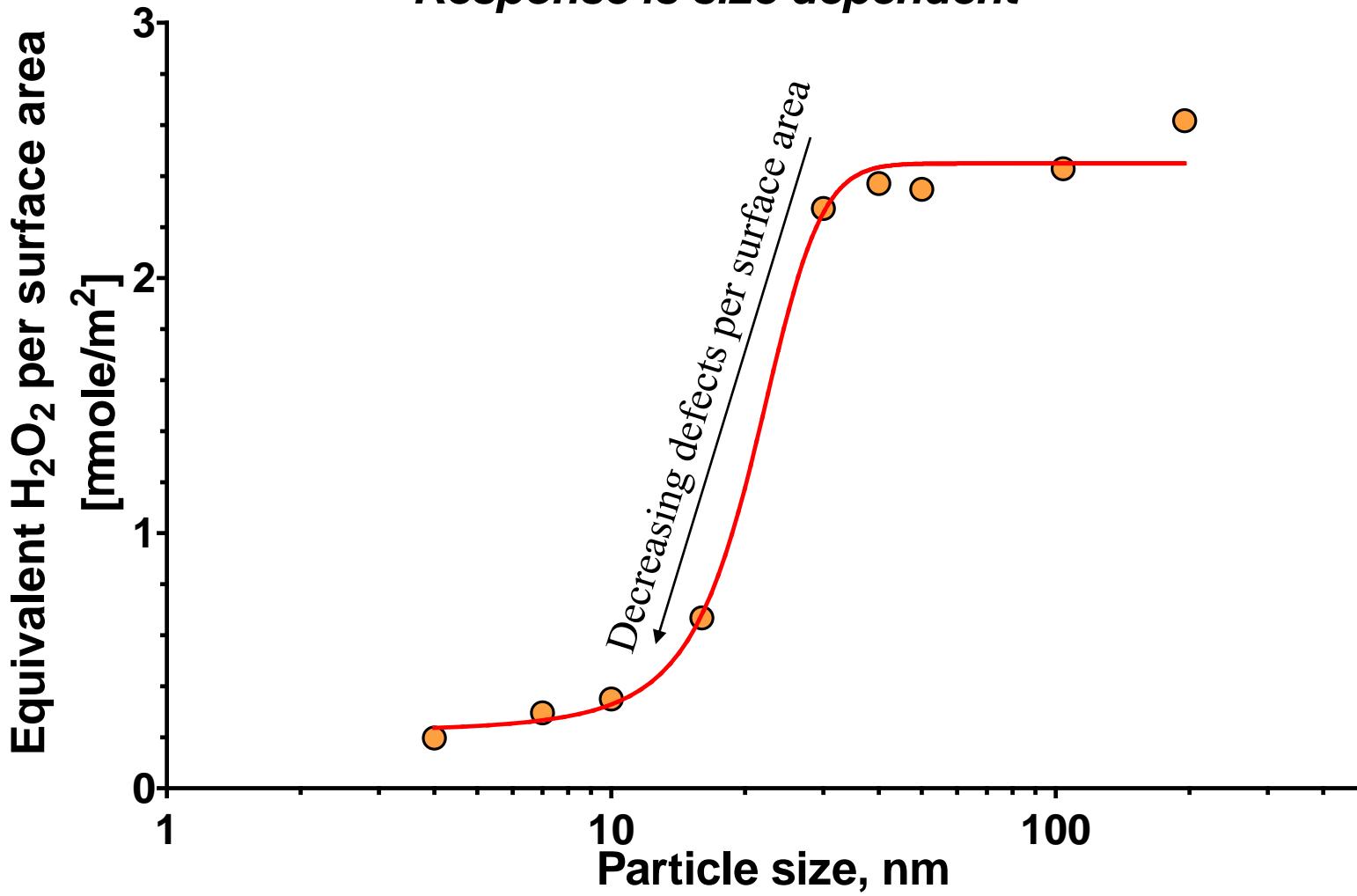
amorphous >

anatase >

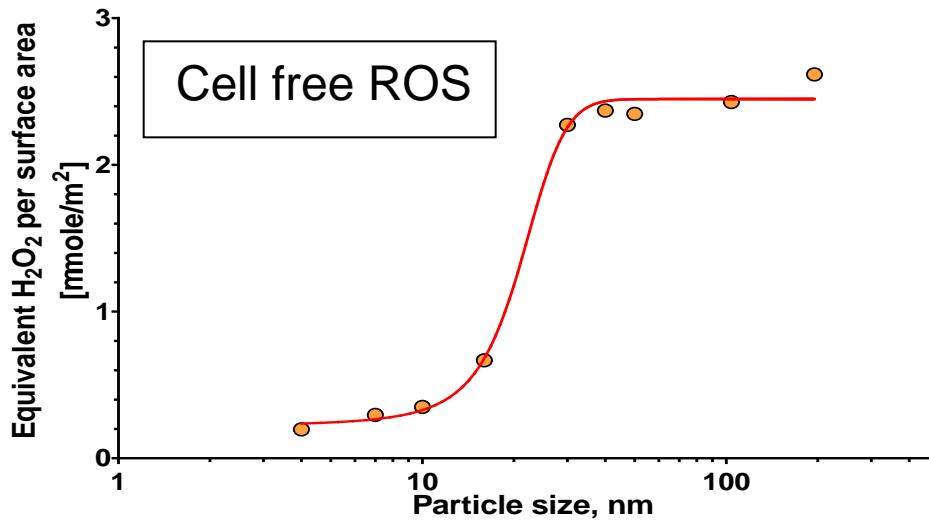
anatase/rutile mixture >

rutile

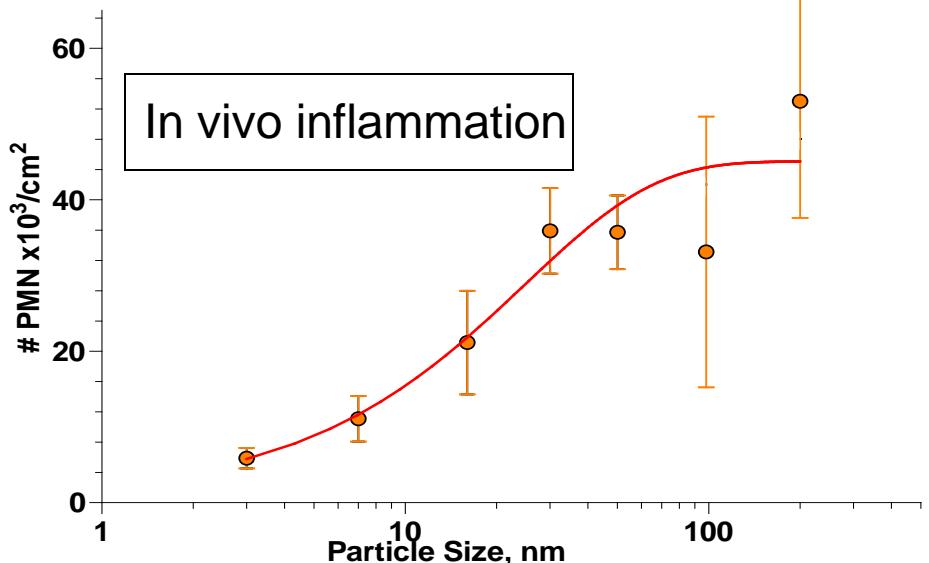
**ROS /cm² Response of Anatase TiO₂ in
Cell Free Assay as a Funtion of Particle Size:
*Response is size dependent***



Maximal ROS /cm² Response to Anatase TiO₂ in Cell Free Assay as a Function of Particle Size

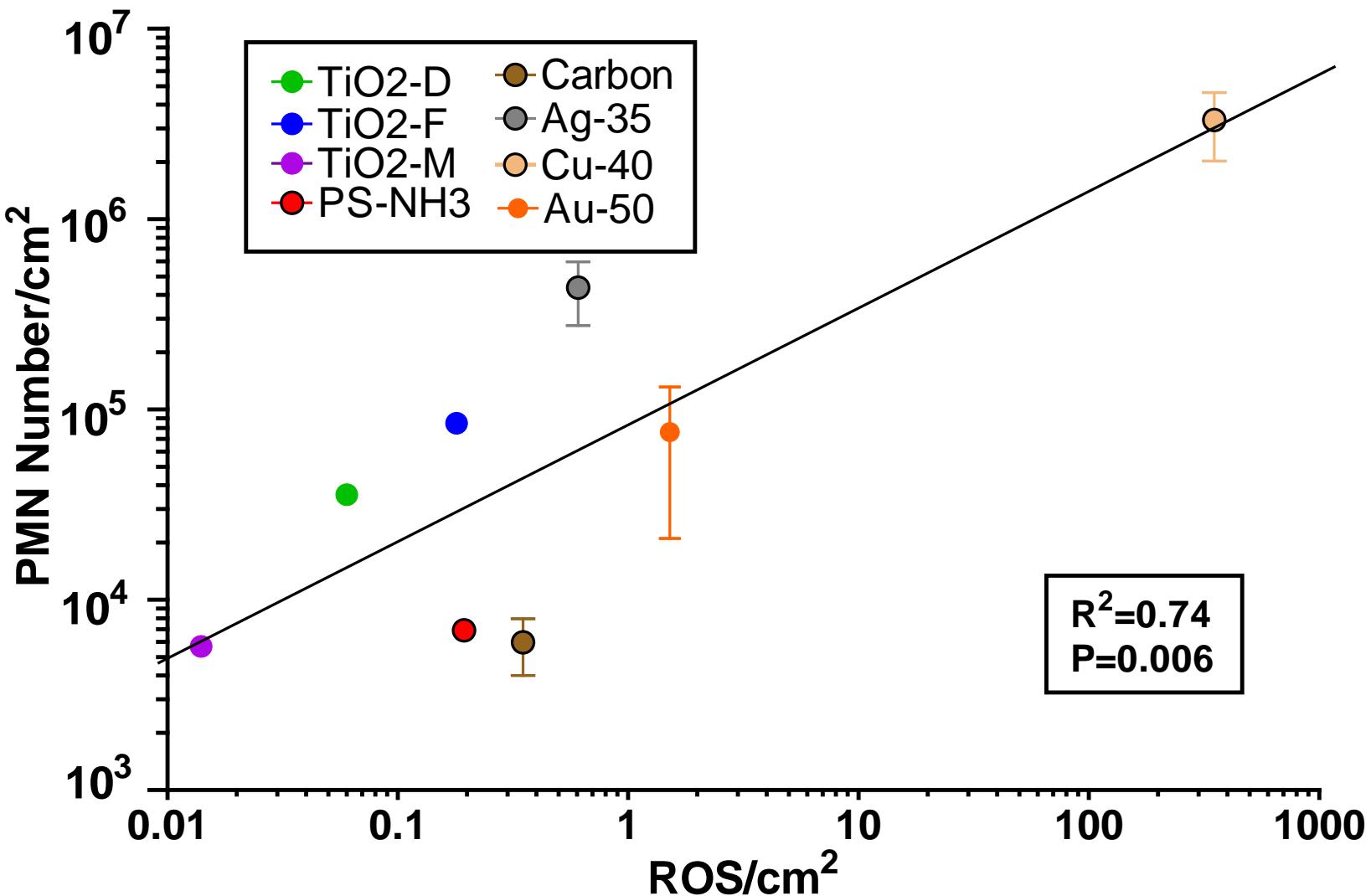


Maximal PMN /cm² Response to Anatase TiO₂ in Rats 24 Hours after Intratrachael Instillation as a Function of Particle Size



**Cell-free ROS (*DCFH oxidation*) Response vs In Vivo Rat PMN (*Intratracheal Instill*)
Response to Nanoparticles
Normalized to Particle Surface Area**

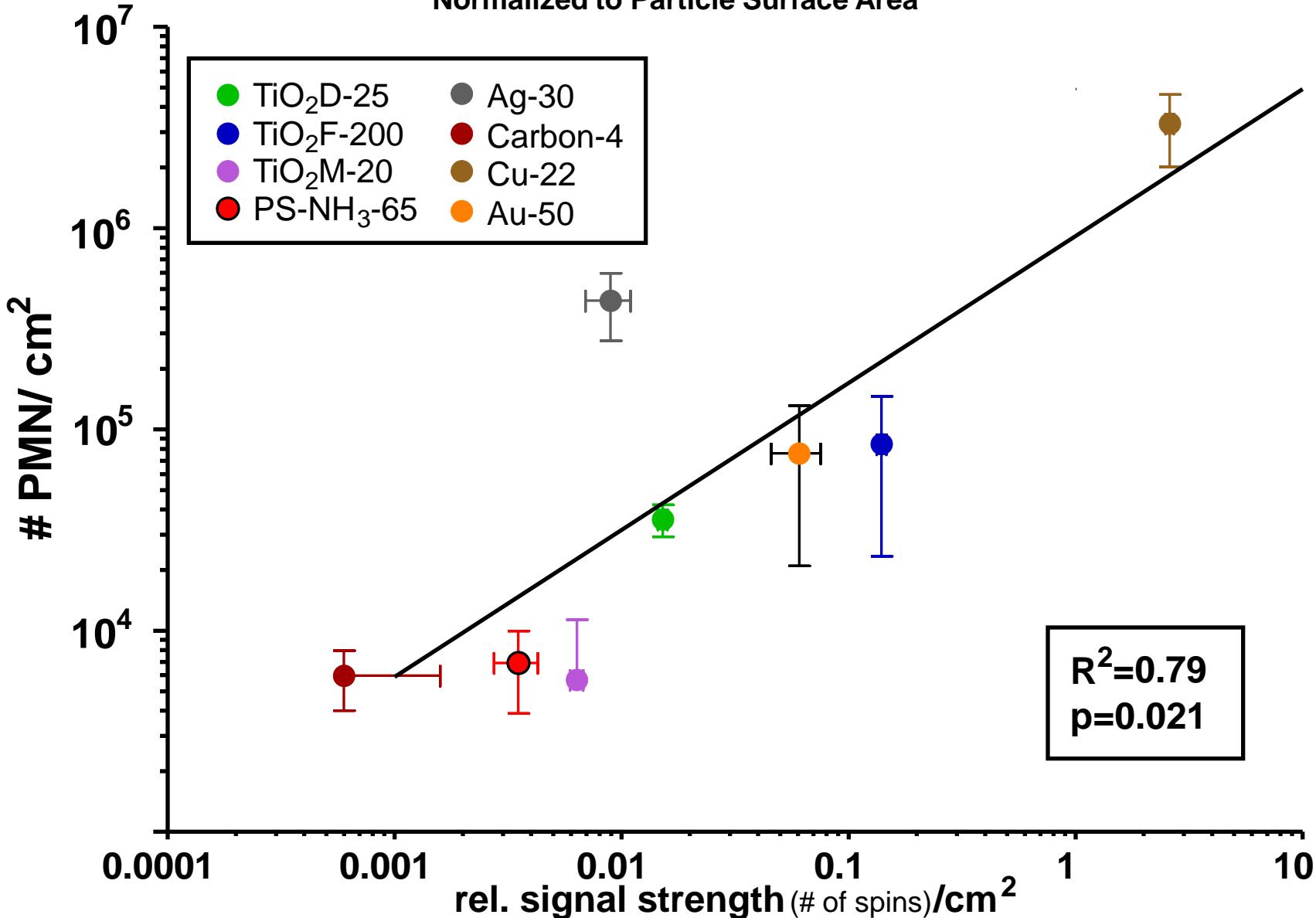
In vivo inflammation



In vitro Cell-free ROS

Rushton et al. 2010

Cell-free ESR (DMPO Spin Trapping) Response vs In Vivo Rat PMN (Intratrach. Instill.)
Response to Nanoparticles
Normalized to Particle Surface Area

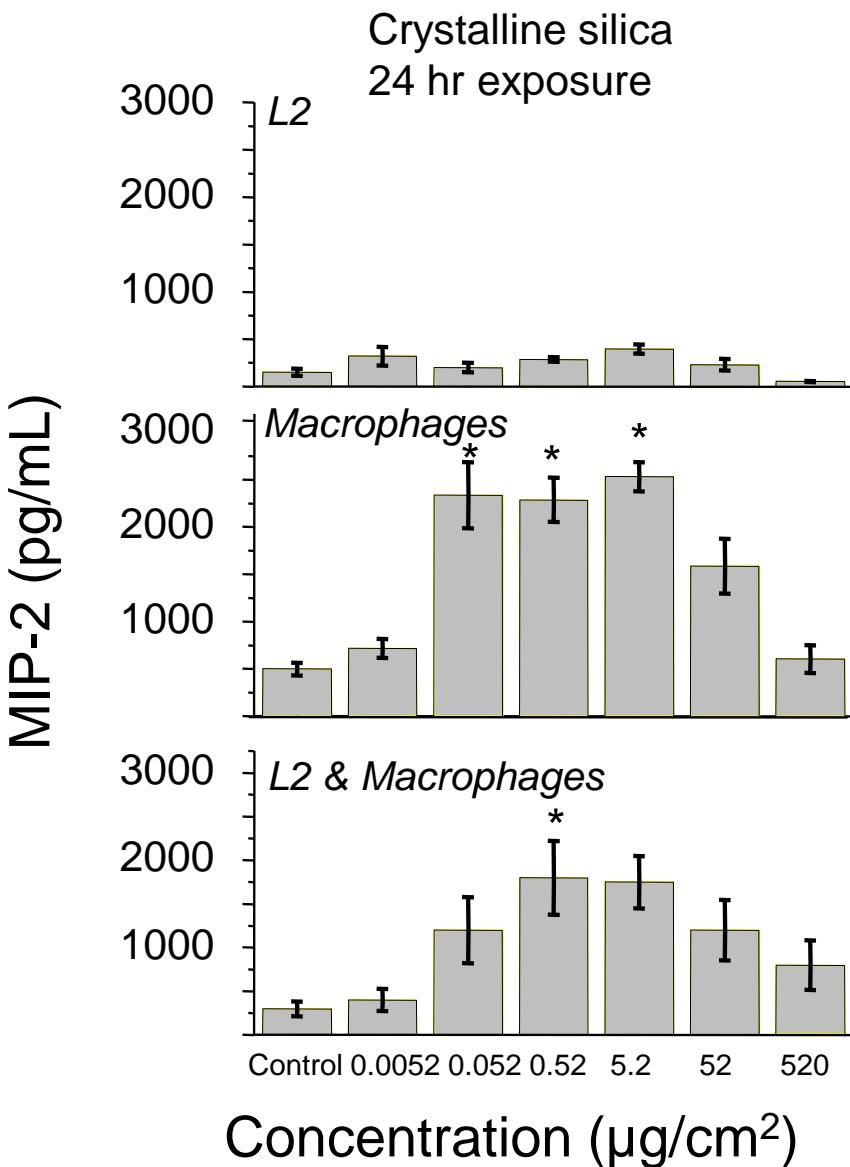
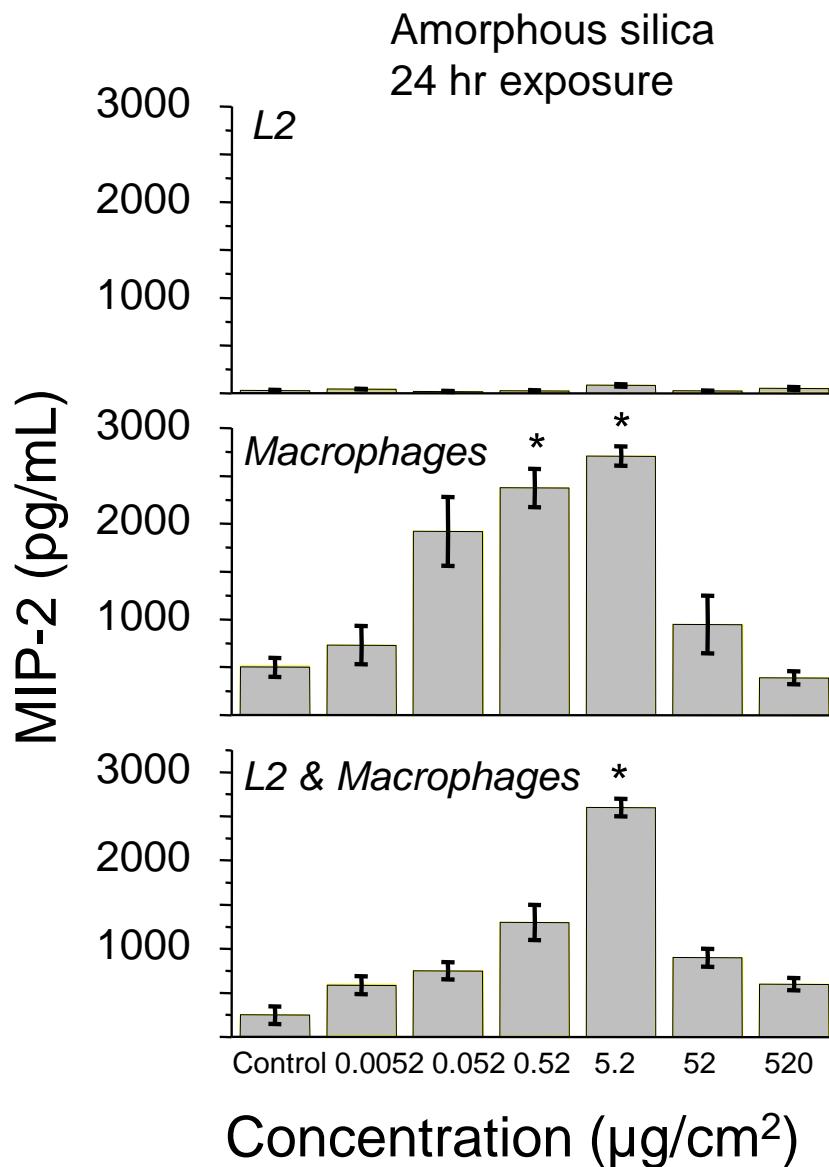


Cell-free ESR

ASSESSING TOXICITY OF FINE AND NANOPARTICLES

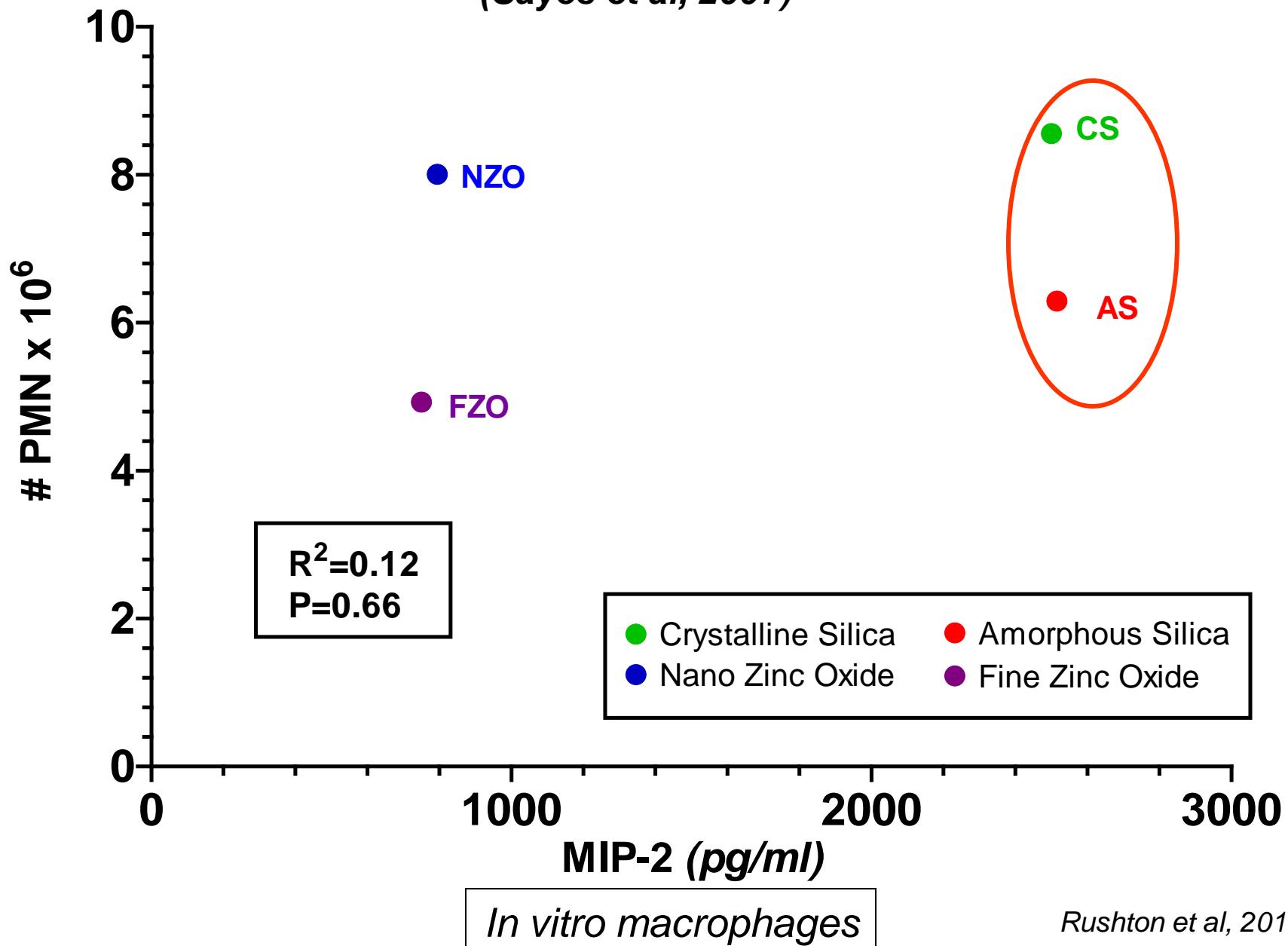
Crystalline Silica; Amorphous Silica; Nano Zinc Oxide; Fine Zinc Oxide

in vitro system, lung epithelial cells (Sayes et al., 2007)

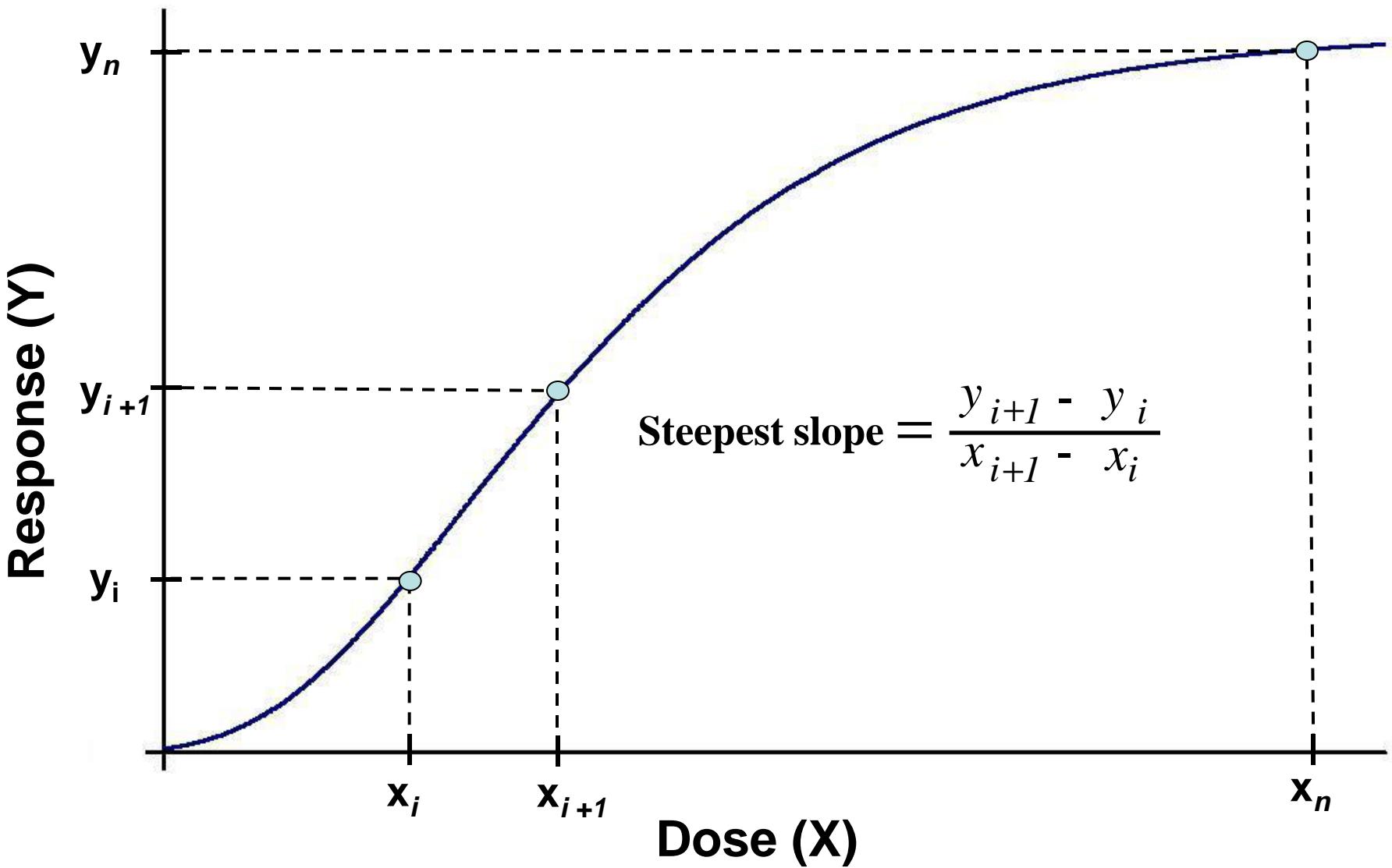


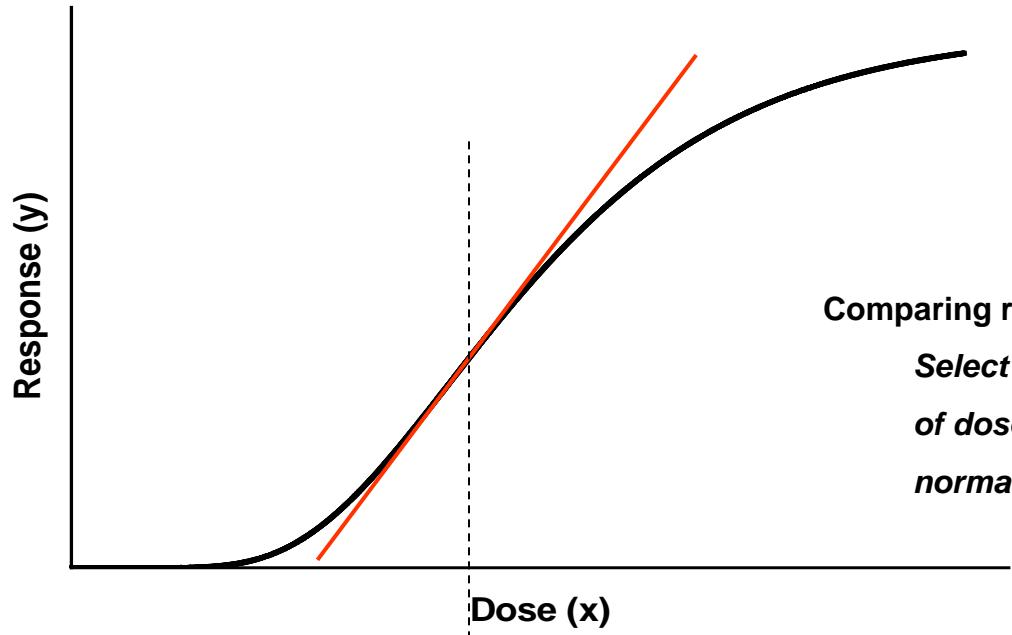
In Vivo/In Vitro Correlation, Highest Measured Responses (Sayes et al, 2007)

In vivo inflammation



Slope (response per unit dose) is Dose Dependent



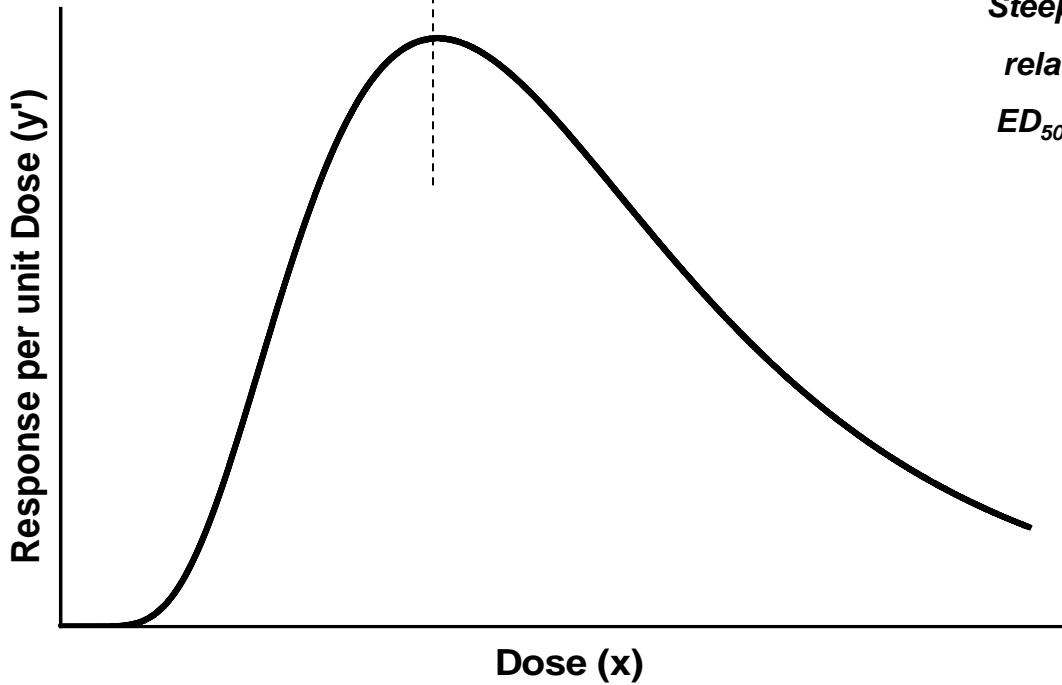


Comparing responses from different assays:

*Select response at corresponding points
of dose response curves of assays,
normalized to a unit of dose.*

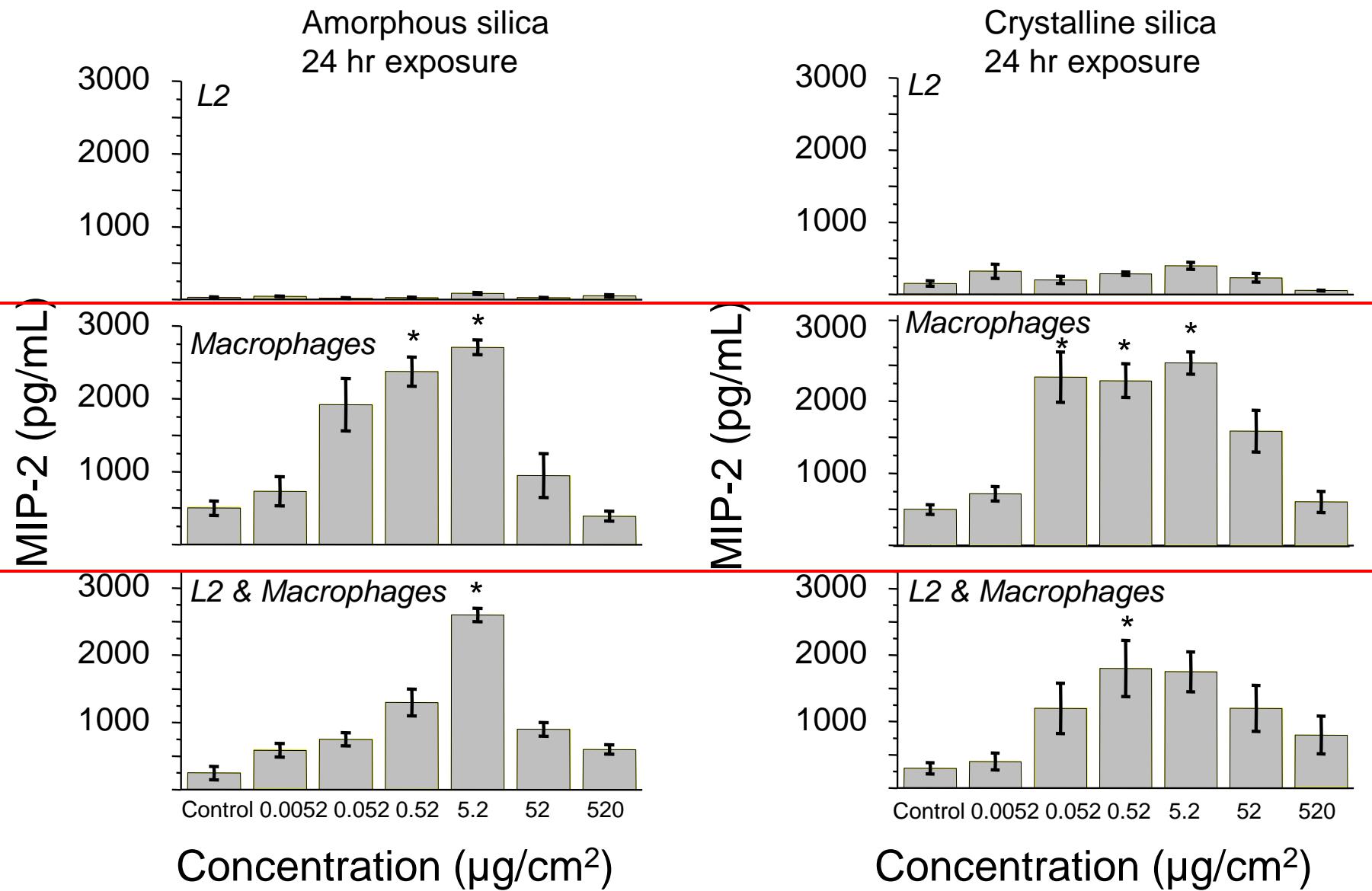
Possibilities:

*Steepest slope of linear dose response
relationship (Max of derivative)
 ED_{50} equivalent of log dose response*

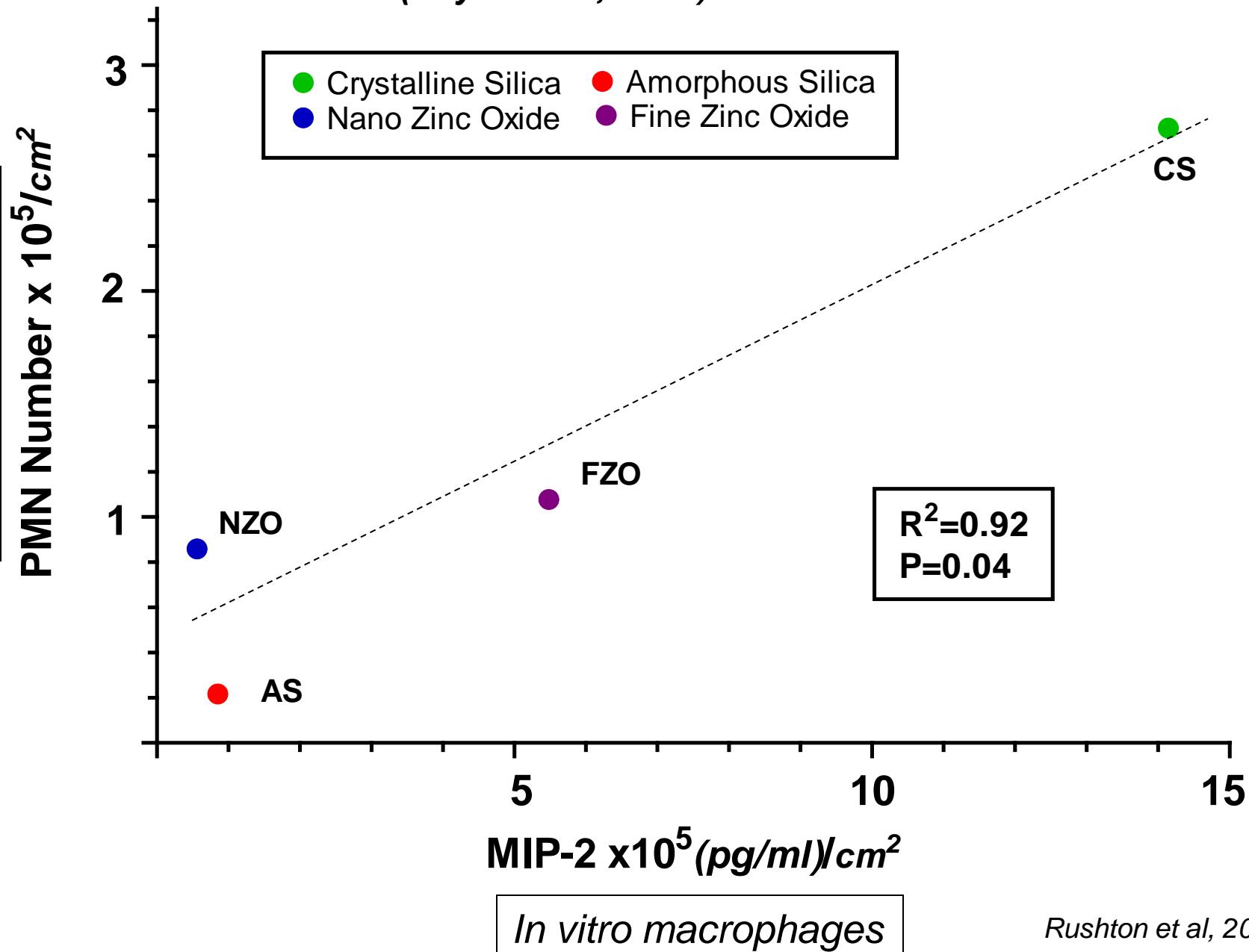


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Crystalline Silica; Amorphous Silica; Nano Zinc Oxide; Fine Zinc Oxide
in vitro system, lung epithelial cells (Sayes et al., 2007)



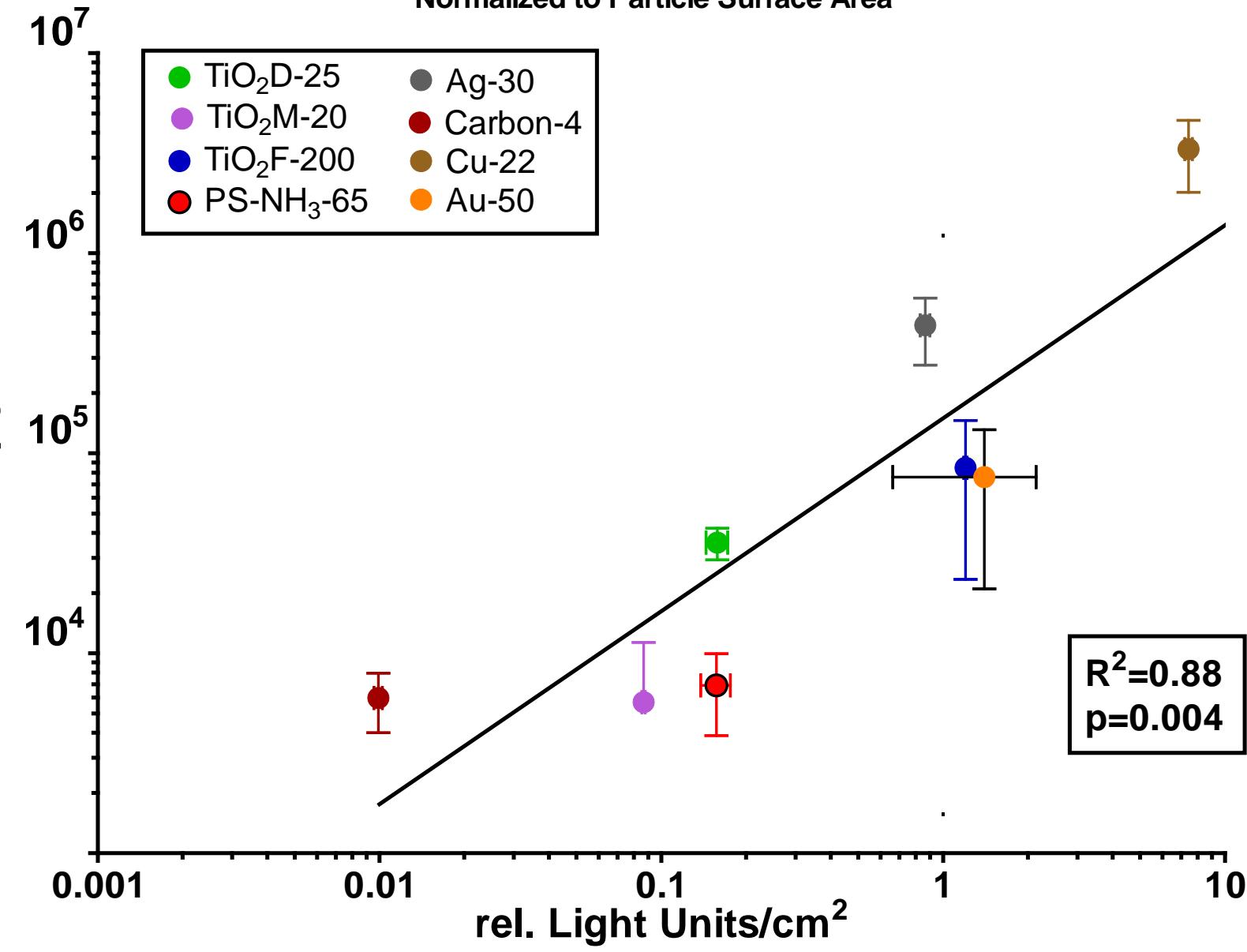
In Vivo/In Vitro Correlation, Highest Responses Per NP Surface Area (Sayes et al, 2007)



Rushton et al, 2010

In vitro (A549Luc1 cells) Luciferase vs *In Vivo* Rat PMN (*Intratrach. Instill.*)
Response to Nanoparticles
Normalized to Particle Surface Area

In vivo inflammation



In vitro A549 cells

Rushton et al, 2010

**Example of categorizing NPs by a hazard scale or
by Reference Particle Equivalent**

(based on maximum effect per NP surface, mass or number,
as derived from dose-response curves)

NP-TYPE	SIZE	HAZARD CATEGORY
Carbon black	41 nm (<i>aggregated</i>)	Very low
TiO ₂ (anat.)	20 nm (<i>aggregated</i>)	
Polystyrene	60 nm (positive charge)	
TiO ₂ (anat./rut.)	25 nm (<i>agggreg/agglom</i>)	Low
Au	50 nm	
Ag	35 nm (<i>agggreg/agglom</i>)	High
Cu	40 nm (<i>agggreg/agglom</i>)	Very high

Example is based on pulmonary inflammatory response in rats (elicited PMN/cm²).
Other *in vitro* or *in vivo* endpoints can be selected, e.g., ROS/cm²; LDH/cm²; MN/cm²;
Prot.Aggr./cm² ...

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			per cm ² (Carbon Black = 1)
Carbon black	41 nm (<i>aggregated</i>)	Very low	1
TiO ₂ (anat.)	20 nm (<i>aggregated</i>)		5
Polystyrene	60 nm (<i>positive charge</i>)		9
TiO ₂ (anat./rut.)	25 nm (<i>aggreg/agglom</i>)	Low	11
Au	50 nm		21
Ag	35 nm (<i>aggreg/agglom</i>)	High	80
Cu	40 nm (<i>aggreg/agglom</i>)	Very high	620

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			per cm ² (Carbon Black = 1)	per gram	per 1.33 x 10 ¹⁵ particles
Carbon black	41 nm (<i>aggregated</i>)	Very low	1	6.1	19.3
TiO ₂ (anat.)	20 nm (<i>aggregated</i>)		5	3.4	2.6
Polystyrene	60 nm (<i>positive charge</i>)		9	6.3	1
TiO ₂ (anat./rut.)	25 nm (<i>aggreg/agglom</i>)	Low	11	5.0	4.2
Au	50 nm		21	1.0	2.1
Ag	35 nm (<i>aggreg/agglom</i>)	High	80	13.3	12.2
Cu	40 nm (<i>aggreg/agglom</i>)	Very high	620	151	508

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Prot.Aggr./cm² ...

Comparing Pulmonary Response to TiO₂ NPs in Rats with same dose administered by Instillation or Inhalation:

- *bolus type vs aerosol delivery*
- *high vs. low dose rate*
- “*prepared*” vs. “*pristine*” NPs
- *more central vs. more peripheral lung deposition*

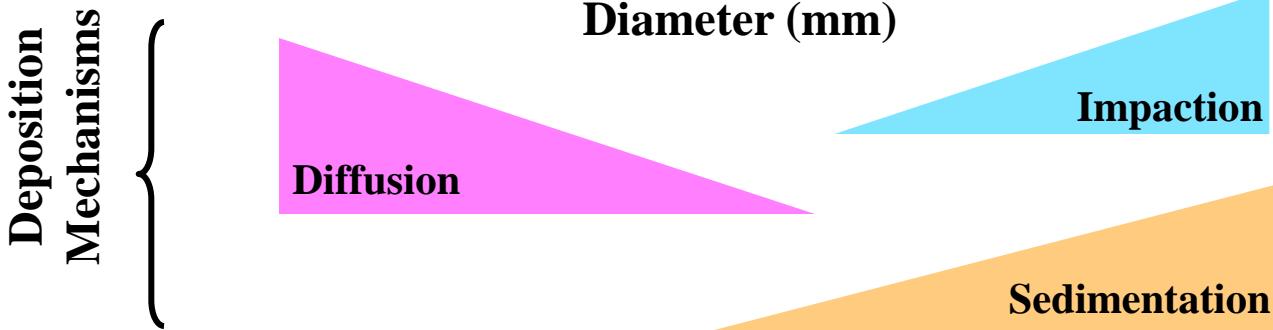
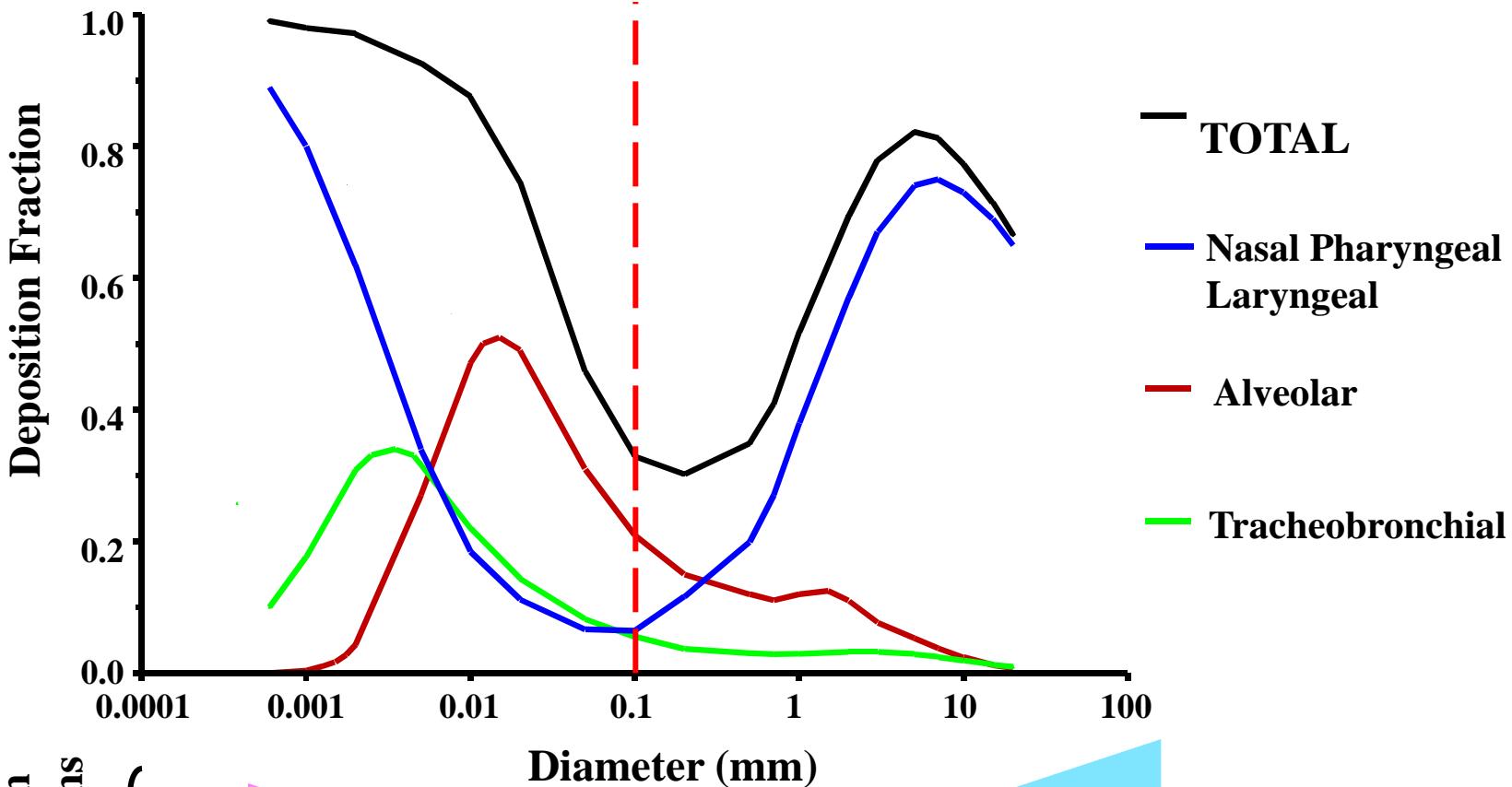
Dispersion as essential step when preparing NPs for toxicity testing using bolus type delivery in vitro and in vivo:

But:

Does it affect NP properties?

Which dispersant?

Which sonication system, time and power?



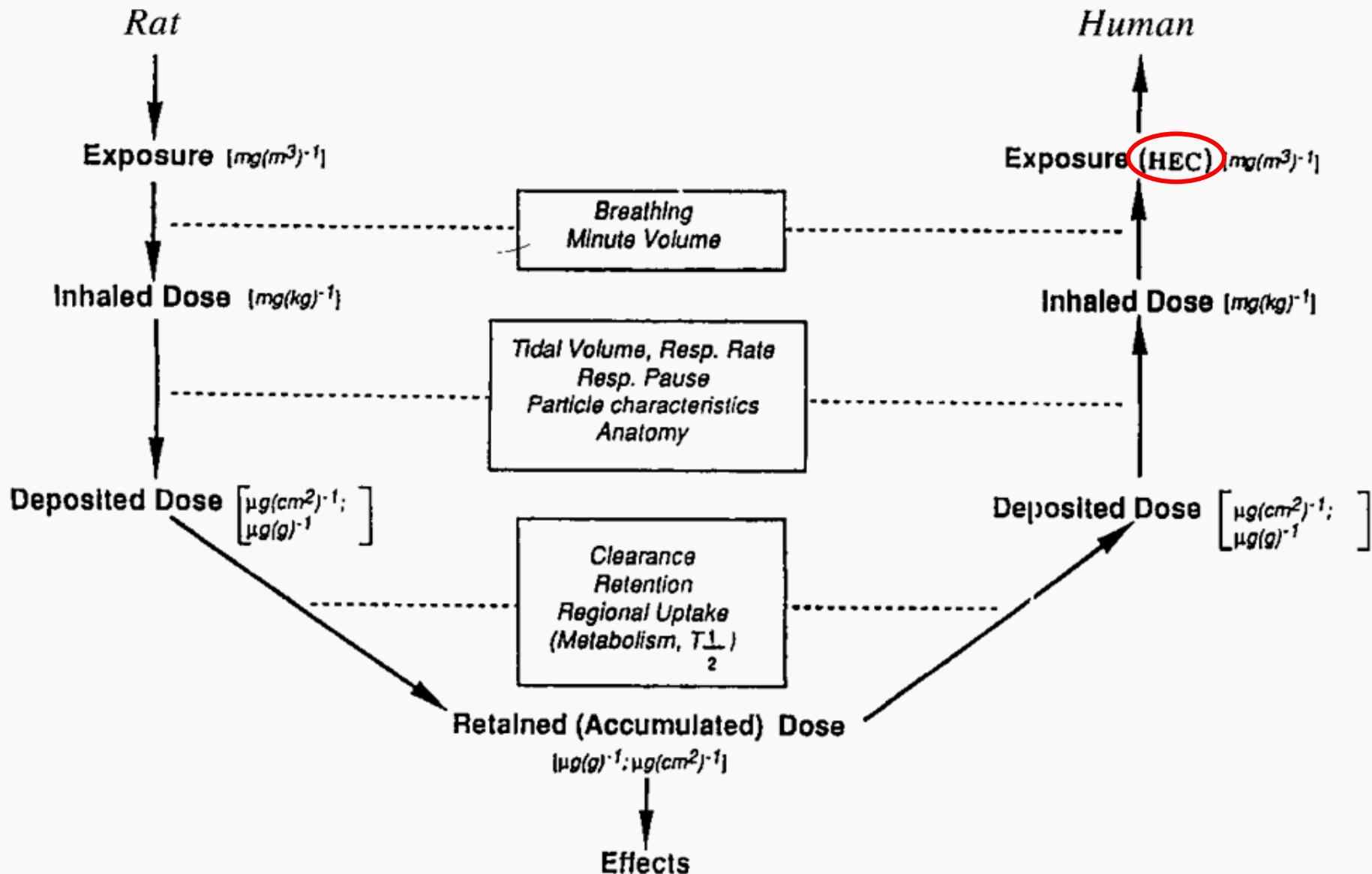
**Deposition per Unit Surface Area (cm^2) over 8 Hour Exposure
at 100 mg/m^3 (nasal breathing, resting conditions)**

Nanoparticle Size: CMD = 20 nm; GSD = 1.0

Density: $\rho = 1 \text{ g/cm}^3$



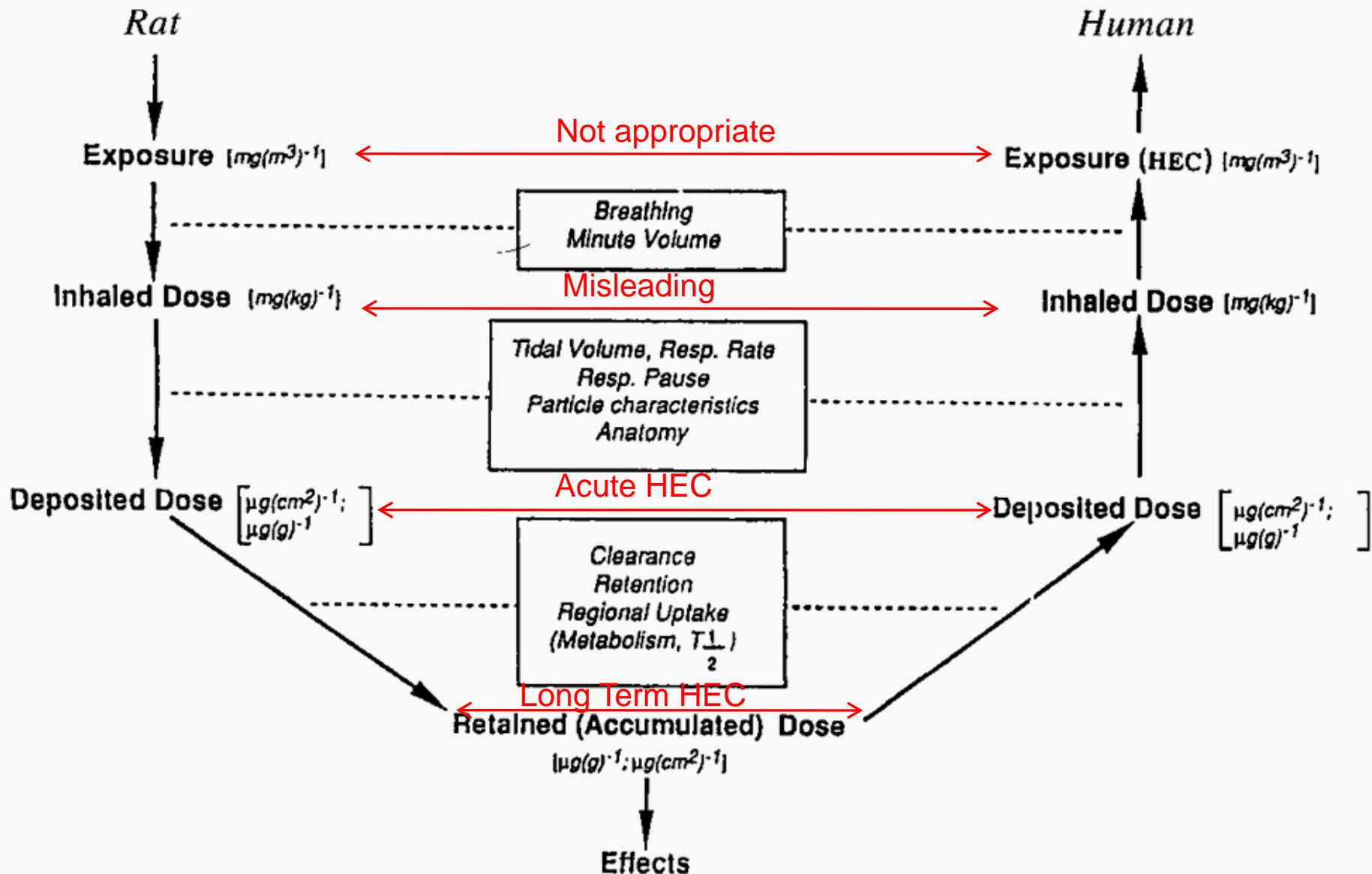
Dosimetric Extrapolation of Inhaled Particles from Rats to Humans



Assumption: If retained dose is the same in rats and humans, then effects will be the same

Oberdörster, 1989

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$$Risk = f(hazard; exposure)$$

